



HAL
open science

Biological activities of triterpenoids from *Poraqueiba sericea* stems

Ilhem Zebiri, Mohamed Haddad, Laurent Duca, Michel Sauvain, Lucie Paloque, Billy Cabanillas, Elsa Rengifo, Jean-Bernard Behr, Laurence Voutquenne-Nazabadioko

► **To cite this version:**

Ilhem Zebiri, Mohamed Haddad, Laurent Duca, Michel Sauvain, Lucie Paloque, et al.. Biological activities of triterpenoids from *Poraqueiba sericea* stems. *Natural Product Research*, 2017, 31 (11), pp.1333-1338. 10.1080/14786419.2016.1241998 . ird-01382564

HAL Id: ird-01382564

<https://ird.hal.science/ird-01382564>

Submitted on 17 Oct 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

SHORT COMMUNICATION

Biological activities of triterpenoids from *Poraqueiba sericea* stems

Ilhem Zebiri^a, Mohamed Haddad^b, Laurent Duca^c, Michel Sauvain^b, Lucie Paloque^{d,e},
Billy Cabanillas^f, Elsa Rengifo^f, Jean-Bernard Behr^a and Laurence Voutquenne-
Nazabadioko^a

^aInstitut de Chimie Moléculaire de Reims UMR 7312 CNRS, Université de Reims, Reims, France; ^bUMR 152 Pharma Dev, Université de Toulouse, IRD, UPS, France; ^cUnité Matrice Extracellulaire et Dynamique Cellulaire (MEDyC), UMR CNRS 7369, Université de Reims, Reims, France; ^dLCC (Laboratoire de Chimie de Coordination), CNRS, Toulouse, France; ^eUniversité de Toulouse, UPS, INPT, Toulouse, France; ^fInstituto de Investigación de la Amazonía Peruana (IIAP), Iquitos, Perú

AQ1

AQ2

ABSTRACT

Eleven compounds were isolated from *Poraqueiba sericea* stems and identified as niga-ichigoside-F1 (**1**), trachelosperoside B1 (**2**), 4-epi-niga-ichigoside (**7**), 19 α -hydroxyasiatic acid (**3**), myrianthic acid (**4**), hyptatic acid (**5**), trachelosperogenin B (**6**), arjunolic acid (**8**), and trachelosperogenin E (**9**), secologanoside (**10**) and secoxyloganin (**11**). Compounds **1–11** were tested for their antileishmanial activities against *Leishmania infantum* promastigotes, **1–6** and **8–11** were tested for their cytotoxic activities on fibroblasts, **1–3**, **5–6**, **8–11** were evaluated for their anti-elastase and anti-acetylcholinesterase assays activities by a spectrophotometric method and **1–2**, **5** and **7–10** were tested using bioautography for their β -glucosidase. No antileishmanial activity was detected; compounds **1**, **2** and **11** showed a moderate cytotoxic activity with IC₅₀ 17.7, 20.5 and 10.9 μ g/mL, respectively; compounds **2**, **8**, **9** and **10** gave a percentage of inhibition ranging from 13 to 16% (at 50 μ g/mL) and compounds **1** and **2** showed an inhibition zone on β -glucosidase and anti-acetylcholinesterase assays.

ARTICLE HISTORY

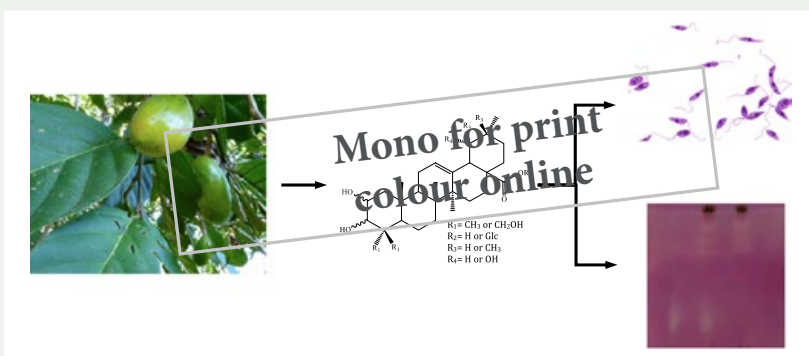

Received 10 May 2016

Accepted 14 September 2016

KEYWORDS

Poraqueiba sericea;
Icacinaeae; triterpenes;
secoiridoids

AQ3

**CONTACT** Mohamed Haddad  mohamed.haddad@ird.fr Supplemental data for this article can be accessed at <http://dx.doi.org/10.1080/14786419.2016.1241998>.

1. Introduction

The genus *Poraqueiba* (Icacinaceae) is composed of three species: *Poraqueiba sericea* Tulp., *Poraqueiba guianensis* Aubl. and *Poraqueiba paraensis* Ducke, large trees growing exclusively in the Amazon. A literature survey showed very few reports on *P. sericea* Tulp, a tree grown initially by Native Americans and propagated by them in Bolivia, Brazil and Peru. This tree is cultivated in Peru for its edible fruit (named Umari in Peru), with a pleasant and a characteristic taste, consumed as **such**, or accompanied by cassava. It is also an ingredient in the soft drink composition called 'Cahuana' with tapioca starch. The oil extracted industrially from the mesocarp is a raw material for this region (food and frying). In industry, the Umari flour replaces wheat flour in the production of adhesives for laminated wood. The only medicinal use concerns the leaves that are used by some tribes as infusion to treat themselves against dysentery (Huamán et al. 2001). Phytochemical investigation of Icacinaceae has resulted in the isolation of indolomonoterpenic alkaloids (Aiyama et al. 1988; Pirillo et al. 1995; Wu et al. 1995, 1996; Srinivas & Das 2003; Khan et al. 2013), flavonoids (Manga et al. 2013), diterpenes (On'okoko et al. 1985), pentacyclic triterpenes (Calderón et al. 2013) and monoterpene iridoids (Braga de Oliveira 1995). The phytochemical study of *P. guianensis* showed the presence of the triterpene icacinic acid, two emmotine derivatives (Braga de Oliveira 1995), one lignan (Goulart et al. 1994) and a secologanoside (Goulart 1983). **Until** now, there is no phytochemical or biological study reported for *Poraqueiba sericea* stems. Indeed, only the oil extracted from the fruit has been the subject of study for the **characterisation** of these fatty acids (Silva 1997).

2. Results and discussion

The structures of the isolated compounds were determined mainly by means of extensive spectroscopic methods including 1D (^1H and ^{13}C NMR), 2D NMR (COSY, ROESY, HSQC, and HMBC) and HR-ESIMS, and by comparison with reported data in the related literature. They are identified as: niga-ichigoside-F1 (**1**) (Bowen-Forbes et al. 2009), trachelosperoside B1 (**2**) (Bowen-Forbes et al. 2009), 19 α -hydroxyasiatic acid (**3**) (Bowen-Forbes et al. 2009), myrianthic acid (**4**) (Hirai et al. 2000), hyptatic acid (**5**) (Bowen-Forbes et al. 2009), trachelosperogenin B (**6**) (Abe & Yamauchi 1987), 4-epi-niga-ichigoside (**7**) and arjunolic acid (**8**) (Bowen-Forbes et al. 2009), trachelosperogenin E (**9**) (Nasser et al. 2006), secologanoside (**10**) and secoxyloganin (**11**) (Calis & Sticher 1984) (Figure 1). Compounds **1–11** were tested for their antileishmanial activity against *Leishmania infantum* promastigotes but no activity was detected (at 100, 10, 1 and 0.1 $\mu\text{g}/\text{mL}$) (Sosa et al. 2016). Compounds **1–6** and **8–11** were also tested (concentration range of 10, 7.5, 5, 2.5 and 1 $\mu\text{g}/\text{mL}$) for their cytotoxicity against human dermal fibroblasts using a MTT assay. Compounds **1**, **2** and **11** showed a very moderate activity with 19.5, 24.6 and 22.4% cell death, respectively, at 10 mg/mL (Table 1) and IC_{50} values of 17.7, 20.5 and 10.9 $\mu\text{g}/\text{mL}$, respectively. The positive control, α -hederin, possessed 71.3% cell death at the same concentration and an IC_{50} of 3.5 $\mu\text{g}/\text{mL}$. Compounds **1–3**, **5–6**, **8–11** were evaluated for their anti-elastase activity and four compounds (**2**, **8–10**) gave a low inhibition percentage ranging from 13 to 16% at 50 $\mu\text{g}/\text{mL}$ (Table 1), twofold less active than ursolic acid used as a standard (31% inhibition) corresponding to IC_{50} values of 24.5, 35.9, 31.0 and 42.0 μM , respectively (10.5 μM for ursolic acid). These compounds should be tested on other elastases because the activities of these enzymes deferred based on their

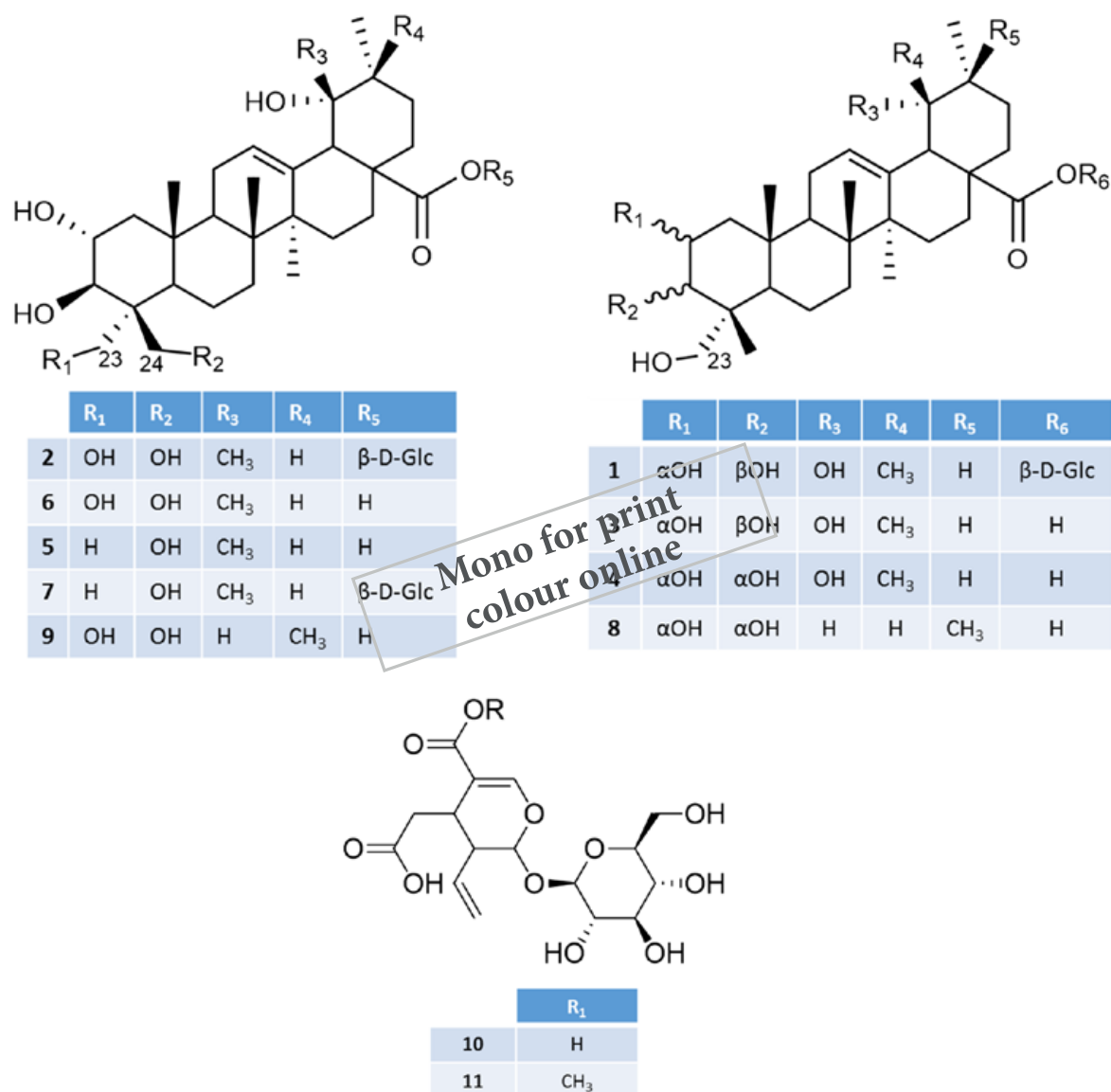


Figure 1. Chemical structures of compounds 1–11.

origin. Therefore, we could have a greater activity on other elastase types. Compounds **1–2**, **5** and **7–10** were tested using bioautography (Camag, Switzerland) for their β-glucosidase and anti-acetylcholinesterase activities at 1 mg/mL (application of 2 μL in 8 mm band length). Only compounds **1** and **2** showed an inhibition zone on the β-glucosidase assay (Substrate: 2-naphthyl-β-glucopyranoside) with an inhibition zone of 11 and 7 mm respectively whereas the positive control, acarbose, showed an inhibition zone of 5 mm only. To verify the detected activity, we used other β and α-glucosidase (0.013 U/mL) tests *in vitro* for compounds **1** and **2** at 500 μg/mL but no activity was revealed by a spectrometrical quantification of the released *p*-nitrophenolate at 410 nm. This suggests that the activity detected on bioautography may be due to an excessive concentration and/or to a reaction of the two saponins with the substrate. The anti-acetylcholinesterase (6.66 U/mL) assay was performed using Fast Blue B salt as reagent, 1-naphthyl acetate as substrate and convallatoxin as positive control. Compounds **1** and **2** showed also an inhibition zone of 0.8 and 0.5 mm, respectively, and 0.4 mm for convallatoxin. Till then we could not verify this activity by *in vitro* tests because we did not have all the required elements.

3. Experimental

See Supplementary material.

4. Conclusion

In this study, three triterpenoids saponins (**1**, **2**, **7**), six triterpenes (**3–6**, **8–9**), and two secoiridoids (**10–11**) were isolated and identified from *P. sericea*. Except for the two secologanositides (**10–11**) (Braga de Oliveira 1995), this is the first report of these triterpenoids in the genus *Poraqueiba* and the Icacinaceae family. Compounds **10** and **11** were considered as chemotaxonomic markers of Icacinaceae species. The isolated compounds were evaluated for several biological activities to valorise our phytochemical study. Compound **1**, **2** and **10** showed moderate cytotoxicity on fibroblasts (IC₅₀: 17.7, 20.5 and 10.9 µg/mL respectively) and compounds **2**, **8**, **9** and **10** showed low antielastase activity with IC₅₀ of 24.5, 35.9, 31.0 and 42.0 µM, respectively. On bioautography, compounds **1** and **2** showed significant inhibition zones for anti-β-glucosidase and anti-acetylcholinesterase activities. Compounds **1–11** did not show any antileishmanial activity. This phytochemical and biological investigation helps us to extend the knowledge about the constituents of the unstudied *Poraqueiba sericea* plant.

Acknowledgements

The authors wish to express thanks to French Minister of Higher Education and Scientific Research for financial support and CNRS, Conseil Regional Champagne Ardenne, Conseil General de la Marne, Ministry of Higher Education and Research (MESR) and EU-programme FEDER to the PIANeT CPER project is gratefully acknowledged.

Disclosure statement

No potential conflict of interest was reported by the authors.

AQ5

References

- Abe F, Yamauchi T. 1987. Trachelosperosides, glycosides of 19α-hydroxyursane-type triterpenoids from *Trachelospermum asiaticum* (Trachelospermum. III). Chem Pharm Bull. 35:1748–1754. AQ6
- Aiyama R, Hisako N, Kenichiro N, Chigiru S, Seigo S. 1988. A camptothecin derivative from *Nothapodytes foetida*. Phytochemistry. 27:3663–3664.
- Bowen-Forbes CS, Mulabagal V, Liu Y, Nair MG. 2009. Ursolic acid analogues: non-phenolic functional food components in Jamaican raspberry fruits. Food Chem. 116:633–637.
- Braga de Oliveira A. 1995. Terpenoids from Amazonian Icacinaceae, Vol. 588. Chemistry of the Amazon, Chapter 10; p. 99–115.
- Calderón AI, Hodel A, Craig E, Gupta MP. 2013. Triterpenes and fatty acids from *Discophora guianensis* identified by GC–MS. Biochem Syst Ecol. 50:16–18.
- Calis I, Sticher O. 1984. Secoiridoid glucosides from *Lonicera periclymenum*. Phytochemistry. 23:2539–2540.
- Goulart MOF. 1983. Estudo químico de quatro espécies vegetais da Amazônia e eletroquímica de emotinas [Ph.D Tesis]. Universidade Federal de Minas Gerais Belo Horizonte. AQ7
- Goulart MOF, Sant'ana AEG, Alves RJ, de Souza Filho JD, Maia JGS, de Oliveira GG, de Oliveira LB. 1994. Icacinic acid, a triterpenoid from *Poraqueiba guianensis*. Phytochemistry. 37:1139–1142.

40

- Hirai N, Sugie M, Wada M, Lahlou EH, Kamo T, Yoshida R, Tsuda M, Ohigashi H. 2000. Triterpene phytoalexins from strawberry fruit. *Biosci Biotechnol Biochem.* 64:1707–1712.
- Huamán PRO, Vargas EBA, Garazatua JF. 2001. Aprovechamiento integral del umarí (*Poraqueiba sericea* Tulasne) en la industria de alimentos. *Rev Amaz Invest Alim.* 1:30–37. AQ8
- 5 Khan N, Tamboli ET, Sharma VK, Kumar S. 2013. Phytochemical and pharmacological aspects of *Nothapodytes nimmoniana*. An overview. *Herba Pol.* 59:53–56.
- Manga A, Gassama A, Sy GY, Bassene E, Lavaud C. 2013. Structural determination of new flavones C-glycosides and trans (S, E)-(–) clovamide isolated from *Icacina senegalensis* Juss leaves (Icacinaceae). *J. Soc. Ouest-Afr. Chim.* 17:15–27.
- 10 Nasser ALM, Mazzolin LP, Hiruma-Lima CA, Santos LS, Eberlin MN, Souza Brito ARM, Vilegas W. 2006. Preparative droplet counter-current chromatography for the separation of the new nor-seco-triterpene and pentacyclic triterpenoids from *Qualea parviflora*. *Chromatographia.* 64:695–699.
- On'okoko P, Vanhaelen M, Vanhaelen-Fastré R, Declercq JP, Van Meerssche M. 1985. The constitution of icacinol, a new diterpene with a pimarane skeleton from *Icacina claessensis*. *Tetrahedron.* 41:745–748.
- 15 Pirillo A, Verotta L, Gariboldi P, Torregiani E, Bombardelli E. 1995. Constituents of *Nothapodytes foetida*. *J Chem Soc Perkin Trans.* 1:583–587.
- Silva NAC. 1997. Characterization of crude, neutro and bleached oil from pulp and peel of jungle fruit *Paraqueiba-sericea*-Tulasne umari. *Grasas Aceites.* 48:11–16.
- 20 Sosa AM, Amaya S, Salamanca Capusiri E, Gilabert M, Bardón A, Giménez A, Vera NR, Borkosky SA. 2016. Active sesquiterpene lactones against *Leishmania amazonensis* and *Leishmania braziliensis*. *Nat Prod Res.* doi: 10.1080/14786419.2015.1126260. AQ9
- Srinivas KVNS, Das B. 2003. 9-Methoxy-20-O-acetylcampthotecin, a minor new alkaloid from *Nothapodytes foetida*. *Biochem Syst Ecol.* 31:85–87.
- 25 Wu TS, Leu YL, Hsu HC, Ou LF, Chen CC, Chen CF, Ou JC, Wu YC. 1995. Constituents and cytotoxic principles of *Nothapodytes foetida*. *Phytochemistry.* 39:383–385.
- Wu TS, Leu YL, Hsu HC, Ou LF, Chen CC, Chen CF, Ou JC, Wu YC. 1996. Nothapodytes A and B from *Nothapodytes foetida*. *Phytochemistry.* 42:907–908.